

## Isolation and Structure of Nodulisporic Acid A<sub>1</sub> and A<sub>2</sub>, Novel Insecticides from a *Nodulisporium Sp.*

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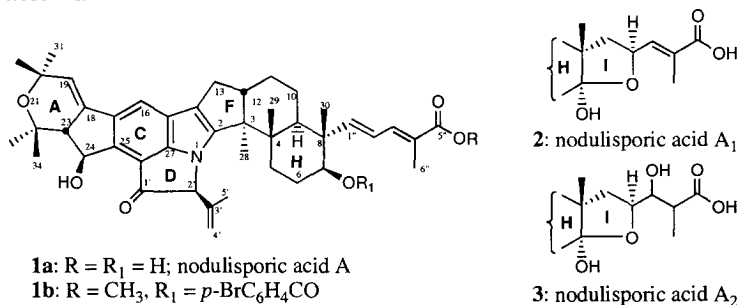
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**Abstract:** The isolation and structure elucidation of two novel indole terpene insecticides nodulisporic acid A<sub>1</sub> (**2**) and A<sub>2</sub> (**3**) from a *Nodulisporium sp.* are reported. © 1999 Elsevier Science Ltd. All rights reserved.

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In our ongoing screening program for biologically active natural products we previously isolated the novel insecticide, nodulisporic acid A (**1a**) from a *Nodulisporium sp.* [1]. Its structure and relative stereochemistry were firmly established on the basis of spectroscopic evidence including the computerized INADEQUATE analysis of Dunkel [2-4] and X-ray diffraction analysis of the *p*-bromobenzoate methyl ester derivative **1b**. The absolute stereochemistry was determined by application of the advanced Mosher method [5]. We now wish to report the isolation and structure determination of two fermentation congeners, nodulisporic acids A<sub>1</sub> (**2**) and A<sub>2</sub> (**3**), which exhibit similar biological profiles to nodulisporic acid A.



Nodulisporic acid A<sub>1</sub> (**2**) has an LD<sub>50</sub> of 0.3 - 1 µg/mL against *Lucilia*, similar to that of **1a** while **3** was slightly less active with an LD<sub>50</sub> of 0.6 - 1.5 µg/mL. Previously, some

structural similarities were noted among several indole diterpenes, such as shearinines and janthitremes, to nodulisporic acid A [1]. Several other indole diterpenes, including paspalines [6], lolitremes [7] and terpendoles [8], have a certain resemblance to A<sub>1</sub> and A.

The methyl ethyl ketone extract of whole broth (pH 5.0) of a 24-day fermentation was evaporated to dryness under vacuum. Nodulisporic acids A<sub>1</sub> and A<sub>2</sub> were isolated from the extract using a silica gel column with 1:1 MeOH:CH<sub>2</sub>Cl<sub>2</sub>. The fractions were dried, chromatographed on a Sephadex LH20 size exclusion column followed by a final purification on a preparative Zorbax C-18 HPLC column (22.5 mm x 250 mm) in 80% CH<sub>3</sub>CN with 0.1% aqueous TFA which gave 1.8 mg of **2**, [ $\alpha$ ]<sup>22</sup>D -10.5 (c 0.6, MeOH) and 1.0 mg of **3**, [ $\alpha$ ]<sup>22</sup>D -21.05 (c 0.38, MeOH).

**Nodulisporic acid A<sub>1</sub>:** High resolution FAB-MS of nodulisporic acid A<sub>1</sub> indicated the molecular formula C<sub>43</sub>H<sub>53</sub>NO<sub>7</sub> (found m/z 695.3806; calcd m/z 695.3822) which was supported by the carbon and carbon-bound proton counts from <sup>13</sup>C NMR and DEPT spectra (Table 1), respectively, assuming three exchangeable protons. The UV spectrum of **2** in MeOH gave absorption bands at  $\lambda_{\max}$  237 ( $\epsilon$  3883), 269 ( $\epsilon$  2767) and 384 ( $\epsilon$  721) nm. The infrared spectrum (ZnSe) showed major absorption bands at  $\nu_{\max}$  3000, 2975, 1710, 1600, 1550, 1470, 1395, 1240, 1090, 1020 and 980 cm<sup>-1</sup>. The <sup>1</sup>H NMR comparison with that of the major component **1a** showed the H7 carbinol resonance missing and modification of the dienoid acid moiety which was confirmed by an extensive number of HMBC correlations indicating the same "western hemisphere" comprising rings A-G in both components (Figure 1). COSY data suggested an ABMX spin system -CH<sub>A</sub>H<sub>B</sub>-CH<sub>M</sub>(O)-CH<sub>X</sub>= [ $J_{AB}$  = 12.6,  $J_{AM}$  = 9.3,  $J_{BM}$  = 7.3,  $J_{MX}$  = 8.8 Hz] which could be expanded by several HMBC correlations to the proposed structure **2**. For example, long-range correlations *via* the two methyl protons (Figure 1) unequivocally positioned the four-spin system with respect to the terminal isobutenoic acid and ring H in **2** and in particular places the hemiketal carbon (106.9 ppm) at C7. This is further corroborated by the three-bond correlations from H1" and H2" to C7 consistent with the presence of the five-membered tetrahydrofuran ring. Substitution of the carbinol group in **1a** by the hemiketal moiety in **2** satisfies the extra oxygen in the molecule and the same number of exchangeable OH protons in both components.

**Nodulisporic acid A<sub>2</sub>:** Nodulisporic acid A<sub>2</sub> was shown to have the molecular weight 713 by ESI-MS (observed at m/z 736 as the [M+Na]<sup>+</sup> ion). High resolution EI-MS indicated the molecular formula C<sub>43</sub>H<sub>53</sub>NO<sub>8</sub> (found m/z 695.3768; calcd m/z 695.3822, [M<sup>+</sup> - H<sub>2</sub>O]) up by a molecule of water from that of nodulisporic acid A<sub>1</sub> (**2**). This was supported by the carbon and carbon-bound proton counts from <sup>13</sup>C NMR and DEPT spectra (Table. 1), respectively, assuming four exchangeable protons. The UV spectrum of **3** in MeOH gave absorption bands at  $\lambda_{\max}$  238 ( $\epsilon$  2740), 267 ( $\epsilon$  2420) and 384 ( $\epsilon$  500) nm. The infrared spectrum (ZnSe) showed major absorption bands at  $\nu_{\max}$  3000, 2975, 1710, 1590, 1470, 1380, 1250, 1090, 1070, 1035 and 1000 cm<sup>-1</sup>. By a similar approach to **2**, structure **3** was proposed for nodulisporic acid A<sub>2</sub>.

**Table 1.**  $^1\text{H}$  and  $^{13}\text{C}$  NMR Assignments of Nodulisporic Acid A<sub>1</sub> (2) and A<sub>2</sub> (3) ( $\text{CD}_2\text{Cl}_2$ , 500 MHz)<sup>a</sup>.

| Carbon | 2                |   | 3                |   |
|--------|------------------|---|------------------|---|
|        | $\delta\text{C}$ | $\delta\text{H}$                              | $\delta\text{C}$ | $\delta\text{H}$                            |
| 2      | 154.8            |   | 154.7            |   |
| 3      | 55.5             |   | 55.4             |   |
| 4      | 39.5             |   | 39.6             |   |
| 5      | 30.0             | ~1.90 m (2H)                                  | 30.7             | ~1.83 m<br>~2.00 m                          |
| 6      | 30.4             | ~1.70 m<br>~1.91 m                            | 30.1             |   |
| 7      | 106.9            |   | 105.5            |   |
| 8      | 49.6             |   | 49.2             |   |
| 9      | 41.3             | 1.68 m  | 47.4             | ~1.70 m                                     |
| 10     | 24.8             | ~1.56 m (2H)                                  | 23.2             | ~1.48 m<br>~1.92 m                          |
| 11     | 26.0             | ~1.9 (2H)                                     | 25.8             | ~1.78 m<br>~1.83 m                          |
| 12     | 48.1             | 2.93 m  | 48.2             | 2.87 m                                      |
| 13     | 27.8             | 2.355 dd (10.8, 14.0)<br>2.755 dd (6.5, 14.0) | 27.8             | 2.33 dd (10.8, 13.9)<br>2.77 dd (6.6, 13.9) |
| 14     | 122.6            |   | 122.5            |   |
| 15     | 121.8            |   | 121.8            |   |
| 16     | 116.7            | 7.72 s  | 116.7            | 7.70 s                                      |
| 17     | 135.9            |   | 135.9            |   |
| 18     | 134.0            |   | 134.0            |   |
| 19     | 122.0            | 6.06 d (2.9)                                  | 122.0            | 6.06 d (2.9)                                |
| 20     | 72.6             |   | 72.6             |   |
| 22     | 73.9             |   | 73.9             |   |
| 23     | 58.2             | 2.81 dd (2.9, 6.3)                            | 58.2             | 2.81 dd (2.9, 6.3)                          |
| 24     | 75.3             | 5.21 d (6.3)                                  | 75.3             | 5.21 d (6.3)                                |
| 25     | 138.3            |   | 138.4            |   |
| 26     | 113.2            |   | 113.2            |   |
| 27     | 162.0            |   | 162.0            |   |
| 28     | 29.9             | 1.34 s  | 29.9             | 1.33 s                                      |
| 29     | 32.0             | 1.31 s  | 32.0             | 1.31 s                                      |
| 30     | 23.4             | 1.122 s                                       | 23.4             | 1.118 s                                     |
| 31     | 30.1             | 1.42 s  | 30.0             | 1.42 s                                      |
| 32     | 15.3             | 0.95 s  | 15.1             | 0.92 s                                      |
| 33     | 16.7             | 1.06 s  | 16.7             | 1.05 s                                      |
| 34     | 17.7             | 1.126 s                                       | 17.6             | 1.112 s                                     |
| 1'     | 198.1            |   | 198.1            |   |
| 2'     | 76.6 br          | 5.10 s  | 76.6             | 5.10 s                                      |
| 3'     | 140.1 br         |   | 140.1            |   |
| 4'     | 117.8 br         | 5.0 br s<br>5.20 dq (~1.2)                    | 117.9            | 5.00 br s<br>5.20 br s                      |
| 5'     | 18.0 br          | 1.42 s  | 17.8             | 1.42 s                                      |
| 1''    | 44.5             | 1.73 dd (9.3, 12.6)<br>2.34 dd (7.3, 12.6)    | 40.3             | 1.48 dd (~9, ~13)<br>1.72 dd (7.5, 12.7)    |
| 2''    | 73.3             | 4.88 br dt (~8)                               | 78.9             | 4.3 m                                       |
| 3''    | 147.1            | 6.93 dq (8.0, 1.3)                            | 81.1             | 3.96 m <sup>b</sup>                         |
| 4''    | 126.8            |   | 40.4             | 2.70 m                                      |
| 5''    | 172.0            |   | 176.3 br         |   |
| 6''    | 12.5             | 1.86 d (1.3)                                  | 13.2             | 1.09 d (7.0)                                |

<sup>a</sup> coupling constants are given in Hz in parentheses. <sup>b</sup> 83.80dd (3.1, 8.3) spiked with DMSO- $d_6$

which makes sense biogenetically as it corresponds to hydration of the side-chain double bond in compound A<sub>1</sub> (2). In particular, the COSY evidence readily showed the contiguous

array of protons  $-\text{CH}_2-\text{CH}(\text{O})-\text{CH}(\text{OH})-\text{CH}(\text{CH}_3)-$  characterizing the side-chain at C8. Again, numerous HMBC correlations corroborated the proposed structure (cf. Figure. 1) especially the three (C3'', C4'', C5'') and four (C7, C8, C9, C1'') long-range correlations from the methyl groups at C4'' and C8, respectively, H4'' to C2'', C3'' and C5'' and H3'' to C1'', C2'', C4'' and C4''-Me.

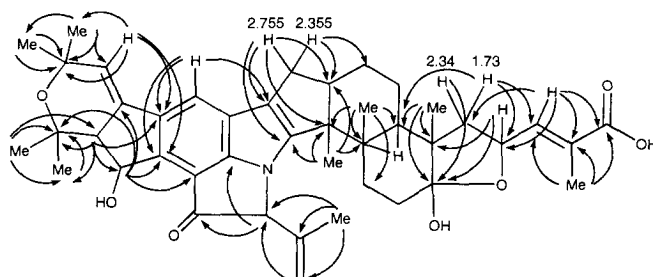


Figure 1: Some HMBC correlations of nodulisporic acid  $A_1$  (**2**)

**Stereochemistry:** Besides many similar NOESY correlations (mixing time 0.5 sec, delay 2.5 sec) indicating the same relative stereochemistry of **1a** and  $A_1$  (**2**), the configuration at C2'' as indicated followed readily from the strong cross peaks between the protons H1'' $\alpha$  and H2'' $\alpha$  on the  $\alpha$ -face of the molecule and between the  $\beta$  oriented methyl group at C8 and the H1'' $\beta$  proton on the  $\beta$ -face. By analogy with **1a**, the complete absolute stereochemistry of **2** is therefore assumed to be the same as in **1a** with the *S* configuration at C2''. Similarly, the vicinal couplings (Table.1) in the tetrahydrofuran ring and NOESY correlations in  $A_2$  by comparison with  $A_1$  suggest the same *S* configuration at C2''.

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#### References and Notes

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